

Austrian, R., and MacLeod, C.M. (1949) J. Exp. Med. 89: 451-460  
Aquisition of M protein by pneumococci through transformation  
reactions.

I - SVI }  
III - A66 fused. { I -  
                  } III - 3M

The "Dawson Rough" seems to correspond to Taylor's ER.

When <sup>or - 36A</sup> II - R36NC } (II; 2'M) was transformed with  
III - A66 TP, III 2'M was obtained.

do, = TPI transformation.

Dawson <sup>ER</sup> Roughs were obtained from R36NC.

Some of these were transformed to III 3M.

from cells which still had some 2'M (<sup>III</sup> 2'M). These may arise  
<sup>obtained to</sup> (subsequently detectable)

This deformation does not take place so regularly. Griffith Roughs  
not tested for TPI.

In vivo: ER + vaccine I  $\frac{1}{10}$   
+ vaccine III  $\frac{2}{10}$

Concurrent acquisition  
of M3 protein noted in  
one case each.

↓  
R  
↓  
II.

Byatt, Pamela H., Jann, G. S. & Salle, A. S. (1948) Variations in pigment production in *Staphylococcus aureus*.

Extracts of chromogenic *S. aureus* (strains?) ~~had~~ transformed white strains to colored. Transformed strains retained bac - character.

Bennett, F.M. + McKie, M. (1929) Type differences amongst  
staphylococcal bacteriophages. *Brit. J. EBMS*. 6: 21-21.

SF: MLE - lac + gel -

Phage B gave three kinds of SF/B: opaque white; colorless &  
translucent; translucent aureo. IB was also resistant to C.  
SF/B was non-lysogenic, but after being kept on agar for some  
weeks gave rise to papillae some of which were of the chalky white  
type, others faintly aureo. Either in this way, or directly  
... SF/B... the aureo-type of SF/B could be obtained.

(1)

Goldstein, Surami (1944) The mechanism of enzyme-inhibitor substrate reactions. J Gen Physiol. 27:529-580

Nm-competitive.

$E$  = total enzyme +  $E_f$   
 $+ =$  free

$$E + I \rightleftharpoons EI. \quad a = \text{fractional activity} = \frac{E_f}{E}$$

$$(1) \quad K_I = \frac{(E_f)(I_f)}{(EI)} = \frac{(E_f)(I - EI)}{(EI)} \quad E = E_f + EI \\ = aE + EI$$

$$(2) \quad I = K_I \frac{(1-a)}{a} + (1-a) E. \quad \text{Let } I' = \frac{I}{K_I}; \quad E_{I'} = \frac{E}{K_I}$$

= "specific concentrations"

$$(3) \quad I' = \frac{1-a}{a} + (1-a) E_{I'} \quad (\text{Zone B}).$$

$$\begin{array}{ll} (\text{free}) & (\text{combined}) \\ \text{Zone A: } I' = \frac{1-a}{a} & (i.e. I \approx I'_+) \\ \text{Zone B: } I' \neq I'_+ \neq EI. & \end{array}$$

$$\text{Zone C: } I' = (1-a) E_{I'} \quad (I' \approx EI)$$



$$a = \frac{v}{V_{\max}} \quad v = k_D(ES) \\ V_{\max} = k_D(E)$$

$$(3B) (4A) \text{ and } S' = \frac{a}{1-a} + a E_S$$

Most enzyme systems operate in zone A., i.e.  $S' = \frac{a}{1-a}$  (MM equation)

They prefer to plot  $\frac{v}{V_{\max}}$  /  $\log_{10} S$ . Consider  $1.1 \times 10^{-3}, 1.25 \times 10^{-3}, 1.7 \times 10^{-3}$  as good fits for  $K_s$ .

The zone B equation is fitted as follows:

$$\frac{S}{a} = K_s \frac{1}{1-a} + E \quad \text{and} \quad \frac{I}{1-a} = K_I \frac{1}{a} + E.$$

$$\frac{V_{max}}{v} = 1 + \left[ K_s + \frac{I}{K_I} \right] \cdot \frac{1}{s}$$

For  $I=0$ ,  $\frac{V_{max}}{v} = 2$  when  $\frac{K_s}{s} = 1$  . ✓

otherwise, for a given, constant activity:

$$\frac{K_s}{s} + \frac{I}{s K_I} = C$$

$$C = \frac{1}{s} K_s + \frac{I}{s} - \cancel{\frac{1}{s}} \cancel{\frac{1}{K_I}}$$

$$SC = K_s + \frac{I}{K_I}$$

$$S_a = 1 + \frac{I}{K_s K_I}$$

$$aS - bI = 1.$$

2)

Competitive equilibrium.

$$\frac{E_f I_f}{(EI)} = K_I \quad \frac{E_f S_f}{(ES)} = K_S$$

$$\frac{(ES)}{E} = a. \quad ES = aE. \quad E = ES + EI + E_f.$$

$$\frac{EI + E_f}{E} = 1 - a \quad EI = (1-a)E - E_f \\ = (1-a)E - \frac{K_S a E}{S - a E}$$

$$I' = \left[ (S' - aE'_S) \left( \frac{1-a}{a} \right) - 1 \right]_{\text{Free}} + \left[ 1 - a \left( 1 + \frac{1}{S' - aE'_S} \right) \right]_{\text{Constrained}} [E'_I] \\ (= (EI)')$$

If  $I_f \approx I$        $I' = (S' - aE'_S) \left( \frac{1-a}{a} \right) - 1$        $GA_I B_S$   
 or if  $EI \approx I$        $I' = \left[ 1 - a \left( 1 + \frac{1}{S' - aE'_S} \right) \right]_{\text{Constrained}} [E'_I]$

He finds  $\frac{I'}{S'} = \frac{1-a}{a}$       i.e. for  $a = \frac{1}{2}$ ,  $\frac{I}{S} = \frac{K_I}{K_S}$ .

$$\frac{1-a}{I'} = \frac{a}{S'}$$

$$\frac{\frac{EI}{E}}{\frac{I'}{I}} = \frac{\frac{ES}{E}}{\frac{S'}{S}} \quad \text{and} \quad \frac{\frac{EI}{I}}{\frac{ES}{S}} = \frac{K_S}{K_I}$$

Hoder, F. + Akano, R., Z. Anatom. 85: 423 - (1935)

Foley, G.E. and Schlesman, H. (1950) <sup>Dec. 1. 1952</sup> ~~J. Exptl.~~

4: 141-149 Some observations on streptococci-dependent strain of *Staphylococcus aureus*. <sup>RR</sup>

Bawden, F.C., Kassam, B., and Nixon, H.L. (1950) The mechanical transmission and seroprepatitis of Rpsvato paracuticle virus.

JGM 4: 210 - 219.

Fleming, A., Verner, A., Kramer, I.R.H., + Hughes, V.H. (1950) The morphology and motility of *Proteus vulgaris* and other organisms re cultured in the presence of penicillin. JGM 4: 257 - 269.

RR

Eriksson, K.R. (1949) Studies on the mode of origin of penicillin resistant staphylococci. Acta path 26: 267 - 279.  
From Univ Inst General Path. Copenhagen.

Broth + various P inoculated with varying amounts ( $10^{-1}$  to  $10^{-6}$ ) of a 24 hr. broth culture. Lately plated loopful (ca. 0.02 ml) on agar. With large inocula, secondary growth is found up to  $\frac{1}{4}$  ou/ml; with initial bacteria of  $10^{-3}$ , no sec. gr., but eventually comes up.  
"Suppose is not correct and that the resistant bacteria appear only after contact with penicillin for some time lengths of time."

Reasoning ?? Notes that with ca  $\frac{1}{8}$  ou/ml and perhaps  $10^{-5}$  ml, any secondary growth was delayed 24-48 hours. In 6 <sup>tubes</sup>, it appeared only after 6 days. "In these cases where the secondary growth appears at such a late period, presumably it can be taken ~~that~~ granted that the growth does not originate from resistant bacteria present in the original culture."

(Some confusion about isolation of pure resistant cultures in test. for stability.)

Found variance in mutant numbers only in 3 ml cultures, not in 15 ml cultures.

Treatment of recombination in tests since 1948

- 1950 Clifton Soltis *et al.* to the bacteria pp 73-75  
"Possibilities of recombination of genes by other than sexual mechanism may exist, and original definition of bacteria as "apparently sex less" organisms is still valid." Fair statement of expts. T+L 1947
- 1949 Burrows *et al.* p. 184 passing reference  
extensive started for general analysis of bacteria T+L 1947.

Stadler, B.A.D. (1949) Measurement of rate of mutation of flagellar genes  
place is ~~not~~ ~~in~~ ~~at~~ ~~in~~ ~~in~~ ~~in~~ ~~in~~ ~~in~~ ~~in~~ J. Hered. 41: 398-413.

[Dept. Genetics & Zoology, Indiana Univ. of Biology, Bloomington, Ind.]

Sloping age + variation in seed, especially older, took by selection.  
occasional mixed strains were found. Some non-viable strains (<2%)  
were found. Some populations at mutational equilibrium, some not.  
Rate of  $3.5 \times 10^{-4}$  / generation found by D. V. Stadler  
at least one. P-405

KR

Klezebeug - Nobel, E. (1947) On the uptake of amino acids and the  
filterability of the  $\text{H}_2\text{O}$ . J. Phys. 11: 95-107.

Uptake filterable, non-filterable, filterability of various amino acids  
as far as possible.

Stern, C. 1936. Somatic crossing over and segregation in *Drosophila melanogaster*  
 Genetics 21: 625-730.

Minute flies ( $M/m$ ) show m spots. Originally interpreted as elimination of  $M$  carrying (deficient chromosome). By use of  $\Theta$ -translocation, it was shown that the  $M$  phenotype (not merely deficiency, covered by duplication) was necessary for spotting. Bobbed ( $bb$ ) spots not found: interpreted as partial elimination.

Autosomal  $M$  also cause X-mosaics (~~or~~  $sn^3$  (segred))  
 However, the  $Bld \rightarrow$  Minute causes X-spots, but not III-spots!!!!

Effect of autosomal  $Mw$  on Notch<sup>8</sup> If. was studied:

$N^8/y$  ♀  $\times$   $sn^3$ ;  $Mw/+$  ♂ Among females:

$N^8/sn^3$  ♀/280       $y/sn^3$   $\underbrace{15/38}$  / No difference.

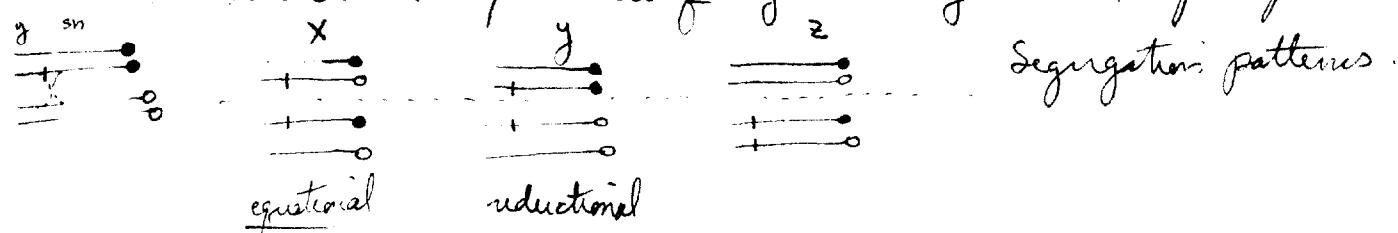
$sn$  setae (elim?  $N^8$ )       $\downarrow$       2  $y$  spots

$2 sn^3$        $\downarrow$       11 twin spots (y) suggesting X-

No y sn spots.  $\therefore$  2-strand and no reduction.

segregation of X-chromosome.

$y sn/++$  flies  $\rightarrow$  110 y sn 43 y  $\overline{7sn}$  spots. y and sn imply somatic crossing-over as well as segregations. But no y-sn twin spots were found, ruling out two-strand crossing over. Complete reduction is ruled out by absence of  $y sn - y - (+)$  triple spots.



Region of crossing-over varies with spot size (developmental stage). Crossing-over to the right of  $sn$  in y sn spots supported by expts. with  $\Theta$  translocation. Segregation is probably nearly always equational.

$bb$  fails to show segregation in + $bb$  flies. Assumption of phenotypic masking seemed unlikely.  $\therefore$  Crossing-over to the right of  $bb$  considered very rare.

Determined X-ploidy of spots by color of 5-6th abd. segments.  
Most spots in females were XX by color.

### Autosomal mosaics

Under influence of autosomal M.

## Secondary Sources:

- (1) Sorsby "Clinical Genetics"; pp/ 337-40; 313-15  
 (2) Kallmann and Sander 1947. in Hoch & Knight, "Epilepsy". Chap. 3  
 (3) Neel 1947 Medicine 26:115. at 123-125

Acc. (3): 25-30% of propositi have family history (5-6x as frequent in parents sibs and children of propositi). monozygotic twin correlation 70%. Quotes Lennox extensively on cerebral dysrhythmia. In 24% of families both parents showed dysr. Obvious complexity.

- (2): Examples in animals; also audiogenic seizures. From Conrad: (incidence figures) %

	gen. pop.	childr.	sibs	neph&nieces	dizyg. mtwins	monozyg cotwins
	.3	6.3	4	1.2	3.1	66.6

## concordance in twins:

	diz	monozyg	
idiopath.	4.3	86.3	Thus even sympt. epilepsy has a genetic component. Index twins were restricted to severe hospital cases.
symptoma.	0	12.5	

also found consanguinity correlations with mental deficiency, but not with schizophrenia.

From Lennox: dysrhythmia

general pop	.10
epileptics	.9
par and sibs	.6

in twins, 85% show concordance of encephalo. if monozyg; 5% if dizyg.

- (1) Similar to 2, but emphasizes consanguinity correl. with phychopathy.

Conclusions: inheritance not simple (probably several different mechanisms). Certainly a very large genetic component in severe cases, from Conrad's twin studies. Most frequent suggestion is dominant with low penetrance, but high incidence of dysrhythmia in both parents of propositi (Lennox) suggests recessive factors also.

(Lennox '47 is Res Pub Ass Res nerv ment dis 26:11)

CC: Dr. Javid

1954  
1/2/54  
copied  
MAY 19 1985

## Conjugation in yeast.

Fowell 1951 *Emphycesis dicayosz*: mating of cells gives  from which either haploid or diploid buds may be generated. Took care to remove profusion buds. Paired 250+/- cells; 30 zygotes formed. 50% egg. → only haploid. Other zygotes → only 2n. "An investigation of sporogenesis revealed that nuclear fusion apparently always occurs in zyg. formed by this proc." Renard 1946 also suggest dic. Also discussed by Gärnemann 16, (Buttermond 75. Bot Rev 1940 6:1) Cyt. Morph. J. Fungi 1928.

Winge & Roberts 1948. Unsuccessful crossing: spores may give hybrid cells "before fusing".

W & L figure  spores. But W'35 also shows substantially complete conjugation and diploid buds. . . some variation.

But note analogy of Fowell's dic. i. congoes formation.

Karnada, H. Jbl. Bakst. I 118: 304-16 (1930)

S. paratyphi B + G+ soil bact  $\rightarrow$  frequent antigenic variation  
in *Salmonella*  $\rightarrow$  enteritidis; breslau.

IPB 35:851

- 19 Bennett 1932 Lysogeny. Pelaletten 69. JBact 34:285  
[Andrewes 7. Proc Roy Soc Med 33 Dec 39]  
Kunzler *Physical Rev* 16:129  
18 Bennett *Acta J Expt BM* 6:27)

BB

Delbrück SGP 23:643 Adsorption vs. Ext. lysis  $\rightarrow$  loss of virus.  
22,365 - Temperature ~~same~~ same as for cell division.

Receptors: 63 - Luria + Freudenreich JEM 59:213 ✓

See Bennett 9. AJEBM 15:227

J Immun 76: 281.

(leave out glucose in virus media)

Tryptose 2% glucose .1% NaCl 1% pH 7

AD Hockley:

$\frac{1}{8}$  (1% agar)  
smear

5 ml phage

2 ml 12-24. bacter.  $\underline{10^8}$  / ml

3 hours later; 5 ml mixture + 3.5 ml .7% agar

pour agar plate =

water agar!!!

Freudzel, J. + Z. Szymanowski, (CRSB 117:543 - 546 (1934))

Recherches sur la Paragglutination: Differenciation des antigènes H et O.

They had shown that P. exhibits a different serological specificity from the "agglutinans composite de Schütze". But the R strains do contain an antigen related to the preceding strains.

~~This~~ paragglutinable strains are homogeneous + repeated resolation indicates that the modification is heritable. Only some E. coli are capable of paragg.

coli-typhoid paragglutination.

The P. coli absorb H-antigens from anti-typhoid sera. The original coli does not. anti-H was removed by absorption on Stanley. There was little further agglutinin absorption. However, there was still considerable aggl. of coli. ∴ Paragg. coli has all H antigens, and a fraction of the O of typhi. anti-P. coli serum has a low titer on heated typhi. Typhi phages don't lyse (P) coli.

2. Balet (I. 121:448 - 451 (1931)) Paragglutination des Bact.

Bang mit Typhusserum. —

L. roni - ab.

Using para A and the tyros, P is also obtained with cross-reactivity, but very little  $\approx$  para B.

Could not transform staph. Relates paraglutination to the  
pn. transformations

Smith WE, J Bact 47:417-418 (1944)

Wahlers + Almader JID 65:147-55 (1907)

Appleby, J. C. J. Bact. 38: 641-51 (1939). Cytology and methods  
of reproduction of two cocci and the possible relation of these organisms  
to a spore forming rod.

~~Appleby~~

Cocci appeared in a culture of the bacillus.

11

Ag' Bact Dept., Univ Reading, England

Sex in Bacteria. Literature:

J. Bact. 50

Nuclei - El. 17. c.

(R)

Baylor, M. B., MO Appleman, OH Sears + GL Clark, J. Bact 50: 249-58 (1945)  
Chem. + Agronomy Illinois

Some morphological characteristics of nucleic acid as shown by the  
electron microscope II. [See Soil Sci Soc Am. P. 7: 269-71 1942]

4-5 granules / cell untreated +  $\approx .02\% \text{NaHCO}_3$ ,  $\approx 2\frac{1}{4}$  hrs. Attempts  
at staining w.g. M <sup>15 min.</sup> saline left mottled cells. (several transparencies; corres-  
ponding to nuclei?) After  $\text{NaHCO}_3$  saline did not remove granules  
acetone removed granules. also  $\text{HNO}_3$ ,  $\text{HCl}$

Krausse, G. J Biol 49: 475 - 1945. A study of ... factors... is presented  
of Bony borders.

low pH n.g.

spores are not found until sugar + glycolipids are added  
+ also the nitrogenous compound.

"healthy cells, facing starvation, combine with..."

See:

Greene HC J Biol 35: 261

Wade et al.

Kuaysi, G + Shudd J Back 45: 349-57 (1973)

Small.

The internal structure of cutaneous dentiger.

Apparent <sup>DR</sup> nuclei as. measured in granular form is 5.56  $\mu$  long.  
Most division of cells occurs in 2  $\mu$  wide bands.

R.R. Mellon, J. Bact. 10: 481-501 (1925) Studies in Bacteric Humidity I Observations on a primitive form of sexuality (zygospore formation) in the colo-typhoid group.

*B. coli* (*Nx*) In patient being given antitoxin appeared as filamentous forms & "many very large coccus-like forms were encountered developing from the filaments."

In vitro, peptone-veal- 5% NaCl broth + 1%  $\text{Na}_2\text{glycophosphate}$  at pH 6.8 autoclaved, ~~pot~~ ~~autoclaved~~, filtered + reautoclaved. Pot reautoclaved in autoclave. Single cell isolate inoculated into broth  $37^\circ$  72h. Thymal R.T.; streaked out on Endo. (with Scott - Jif Sep. 8) was incubated at  $37^\circ$  18-24 hours, periphery of colonies was fungoid & zygospore formative.

"no attempt has been made to study the fate of these spore-like bodies".

Similar forms were found in small cells.

No convincing evidence of origin from  $> 1$  cell.

Mystic on sexuality  
& variability  
Do not understand bases  
of relationship.

Assumes that cell-fusion has taken place. Criticizes Hengquist.

"makes it necessary... to rule out the purely symbiotic influence of the accompanying strain".

10: 579-88 (1925)